

Delivery system for pharmaceutical agents

Cross References to Related Applications

5           This application claims the priority of European patent application 02 014 991.0, filed July 10, 2002, the disclosure of which is incorporated herein by reference in its entirety.

10           Technical Field

          The present invention relates to a delivery system for pharmaceutical agents. Said delivery system comprises liposomes which comprise in their internal compartment a pharmaceutical agent to be delivered and which  
15       have linked to their external surface the cell adhesion molecule NCAM or a fragment thereof.

Background Art

20           Liposomes have been widely used for the delivery of a variety of material to cells such as e.g. the-  
          rapeutical drugs and DNA. In order to target the delivery of the encapsulated material to a specific cell type, va-  
25       rious liposome modifications have been developed. In one approach adhesive peptides binding the cell surface mole-  
          cule ICAM-1 (intercellular adhesion molecule-1) on the target cells were linked to the liposome surface (Jaafari  
MR, Foldvari M, J. Pharm. Sci. 91(2): 396-404, 2002). In  
30       another approach cationic liposomes were targeted in vi-  
          tro to specific tumor cells by means of monoclonal anti-  
          bodies (Kao GY et al., Cancer Gene Ther. 3(4): 250-6, 1996).

          The hitherto know liposome based delivery sy-  
35       stems have a major drawback, since they do not allow an efficient delivery of pharmaceutical agents to cells.

There is therefore a need for alternative liposome based delivery systems for pharmaceutical agents.

### Disclosure of the Invention

5

Hence it is an object of the present invention to provide a system for the intracellular delivery of pharmaceutical agents wherein said system comprises liposomes which comprise in their internal compartment a  
10 pharmaceutical agent and which have linked to their external surface the cell adhesion molecule NCAM (neural cell adhesion molecule) or a fragment thereof.

In a preferred embodiment of the present invention said fragment of NCAM comprises IG (immunoglobulin)  
15 lin ) loop domains I, II and III.

In a preferred embodiment of the present invention said cell adhesion molecule is linked to said external surface of said liposomes via a transmembrane domain or a hydrophobic anchor molecule which allows the  
20 cell adhesion protein to be tethered to the lipid bilayer such as e.g. a GPI anchor or by isoprenylation of the cell adhesion molecule.

Methods for the production of liposomes are known to the man skilled in the art and e.g. described in  
25 *Liposomes a practical approach*. RRC New Editor. IRL press at oxford university press. 1990.

The term "cell adhesion molecule" as used herein encompasses at least cell adhesion molecules isolated from their natural source, cell adhesion molecules  
30 produced by recombinant technology, and cell adhesion molecules produced by *in vitro* peptide synthesis and fragments thereof.

The term pharmaceutical agent as used herein encompasses all classes of chemical compounds exerting an  
35 effect in a biological system. Preferred pharmaceutical agents for the use in the present invention are molecules selected from the group consisting of DNA, RNA, oligo-

nucleotides, polypeptides, peptides, antineoplastic agents, hormones, vitamins, enzymes, antivirals, antibiotics, antiinflammatories, antiprotozoans, antirheumatics, radioactive compounds, antibodies, prodrugs, and combinations thereof.

In a preferred embodiment of the present invention said pharmaceutical agent is a DNA, preferably a cDNA which is operably linked to a gene expression construct allowing expression of said cDNA, more preferably said cDNA encodes a functional protein. Typically, an expression construct comprises the regulatory sequences required to achieve expression in the host cell and it may contain necessary sequences required for plasmid replication in order to exist in an episomal state, or it may be designed for chromosomal integration. The term regulatory sequence as used herein encompasses both the native regulatory sequence of the gene to be expressed from said expression construct and heterologous regulatory sequences. The construction of such expression constructs are known to the man skilled in the art and are e.g. described in *Sambrook et al., Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Laboratory, 2001.*

When said expression construct is intended for chromosomal integration, the integration into the host genome is preferably achieved by including cis-elements into the expression construct that can be recognized by a transiently expressed transposase such as e.g. Sleeping Beauty. The transposase can be expressed temporarily by including the cognate mRNA into the delivery system. Said expression construct intended for chromosomal integration preferably comprises cis-elements preventing transcriptional silencing that follows random integration.

In a much preferred embodiment said DNA construct is designed for selective integration into the genome of host cells. For this purpose, the integrase of phi bacteriophage C31 is preferably used (Ortiz-Urda S.

et al., Nat Med 2002, 8 (10): 1166-70). When using the phi bacteriophage C31 integrase the DNA construct comprises the AttB sequence which is recognized by C31 integrase. The integrase can e.g. be expressed in the host cells  
5 from a adenoviral expression vector which is delivered into the cells with the inventive delivery system. It is as well possible to include into the inventive delivery system *in vitro* produced mRNA coding for C31 integrase. The transient expression of C31 integrase in host cells  
10 via a recombinant-adenovirus or via a mRNA bears on the notion that long term expression of such DNA rearranging enzymes could itself be genotoxic and therefore should be avoided.

In a another preferred embodiment said DNA  
15 construct further comprises locus control elements (LCR) e.g. matrix attachment sites.

In a much preferred embodiment of the present invention said DNA encodes the human dystrophin protein.

In a preferred embodiment of the invention  
20 said delivery system comprising as pharmaceutical agent a DNA further comprises a DNA compacting agent such as e.g. cationic peptides. Said DNA compacting agents reduce the size of the liposome particles. Preferred DNA compacting agents are reversibly cross-linkable cations that  
25 can be crosslinked after particle formation. A much preferred crosslink is a thio bridge.

In another preferred embodiment of the invention said delivery system comprising as pharmaceutical agent a DNA further comprises a chemical inclusion such  
30 as e.g. secondary and tertiary amines and/or a biological inclusion such as e.g. adeno fiber, to breach the intracellular endosomal barrier.

Said delivery system comprising as pharmaceutical agent a DNA preferably comprises nuclear localisation signals, more preferably said nuclear localisation signals are PNA linked peptides and/or PNA linked ligands. Said PNA linked peptides or PNA linked ligands can  
35

be picked up by nuclear-cytoplasmic shuttles and lead to the nuclear delivery of the DNA.

In another preferred embodiment of the invention said delivery system comprising as pharmaceutical agent a DNA further comprises an anti-apoptotic activity. Preferred anti-apoptotic activities are selected from the group consisting of Bcl-2, a small interfering RNA (siRNA) directed against Bax and a peptid comprising caspase inhibitor sequences. The preferred anti-apoptotic activity for use with the inventive delivery system is Bcl-XL. The inventive delivery system can comprise one of said anti-apoptotic activities or a mixture thereof. Said activities are included into the inventive delivery system in order to avoid that successfully transfected cells are lost due to apoptosis. The anti-apoptotic activity can e.g. be present in the inventive delivery system in form of an adenoviral expression vector or in form of *in vitro* produced mRNA.

It has to be understood that the inventive delivery system for DNA can comprise any combination of the structural elements disclosed in the above embodiments.

Another object of the present invention is a pharmaceutical composition comprising a delivery system according to the present invention.

The delivery system and the pharmaceutical composition of the present invention are suitable tools for the treatment of various human diseases such as e.g. genetic diseases and cancer as well as for gene therapy. The inventive delivery system comprising as pharmaceutical agent a DNA encoding a functional dystrophin protein is a useful tool for the treatment of Duchenne muscular dystrophy. Depending on the cell adhesion molecule included in the inventive delivery system a pharmaceutical agent can be targeted to specific cells and/or tissue in the human or animal body. The delivery system of the pre-

sent invention can as well be used for the introduction of pharmaceutical agents into cells *in vitro*.

The inventive delivery system or pharmaceutical composition can be administered intravenously as a bolus or by continuous infusion over a period of time, by intramuscular, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. Such pharmaceutical compositions can encompass pharmaceutically acceptable carriers that are inherently nontoxic and nontherapeutic. Examples of such carriers include ion exchangers, alumina, aluminum stearate, lecithin, serum proteins, such as human serum albumin, buffers such as phosphate or glycine, sorbic acid, potassium sorbate, partial glyceride mixtures of saturated vegetable fatty acids, water, salts, or electrolytes such as protamine sulfate, sodium chloride, metal salts, colloidal silica, magnesium trisilicate, polyvinyl pyrrolidone, cellulosic polymers, and polyethylene glycol.

The invention is now further illustrated by means of examples.

Example 1: Production of liposomes having incorporated into their membrane cell adhesion molecules

Dry 0.5  $\mu\text{mol}$  of dioleoyl phosphatidyl choline under nitrogen in a disposable glass tube. Evacuate in dessicator under vacuum for 30 minutes. Add buffer/dH<sub>2</sub>O to required volume and scrape the sides of the glass tube to dislodge the lipid. Add protein a  $1\mu\text{g}/\mu\text{l}$  of lipid used. Vortex for 30 seconds. Sonicate twice in a bath sonicator at 7 degree for 15 seconds.

This procedure results in multilamellar vesicles that become small unilamellar vesicles (SUV) with prolonged sonication time. To make large unilamellar vesicles, use the extruder.

Example 2: Expression of lipid tagged proteins and preparation of liposomes having incorporated said lipid tagged proteins.

5                   E. coli strain HB101 was transformed with a plasmid and cultured in LB medium with ampicillin at 37°C. The plasmid encodes a hexahistidiny1 tail for purification of the tagged protein. The DNA sequence encoding the signal peptide and the N-terminal amino acid residues of the major lipoprotein of E. coli (lpp) was linked to the  
10 DNA encoding the protein. The N-terminal cysteine residue of the resulting protein is enzymatically linked with the lipids in the bacterial membrane. After induction with IPTG, the cells were cultured another 12 h at 30°C and  
15 harvested by centrifugation. The cells from 11 culture were suspended in 50 ml lysis buffer and lysed by sonication. The cell envelopes were collected by ultracentrifugation (150'000 g, 1h, 4°C), the pellet was suspended in HEPES buffer (pH 7.4) containing 1% (w/v) Triton-X-100.  
20 The sample was applied to a Ni<sup>2+</sup> column to purify the lipid tagged proteins having a hexahistidiny1 tail.

Ten milligrams of phosphatidylcholine was dissolved in 1 ml of chloroform in a test tube. After drying well under a stream of nitrogen, it was suspended  
25 in 1 ml HEPES buffer solution (pH 7.4) and sonicated for 10 min. After centrifugation at 30'000 g for 20 min, the final pellet of liposomes was suspended in 1 ml of HEPES buffer. Then the solution of purified lipid tagged proteins were added to the resulting liposomes with stirring  
30 at 4°C.

While there are shown and described presently preferred embodiments of the invention, it is to be distinctly understood that the invention is not limited thereto but may be otherwise variously embodied and practiced within the scope of the following claims.  
35